



Even Nuclear Pores Get Old; But How?

Aisha Hamilton Frøiland, Evgeny Onishchenko

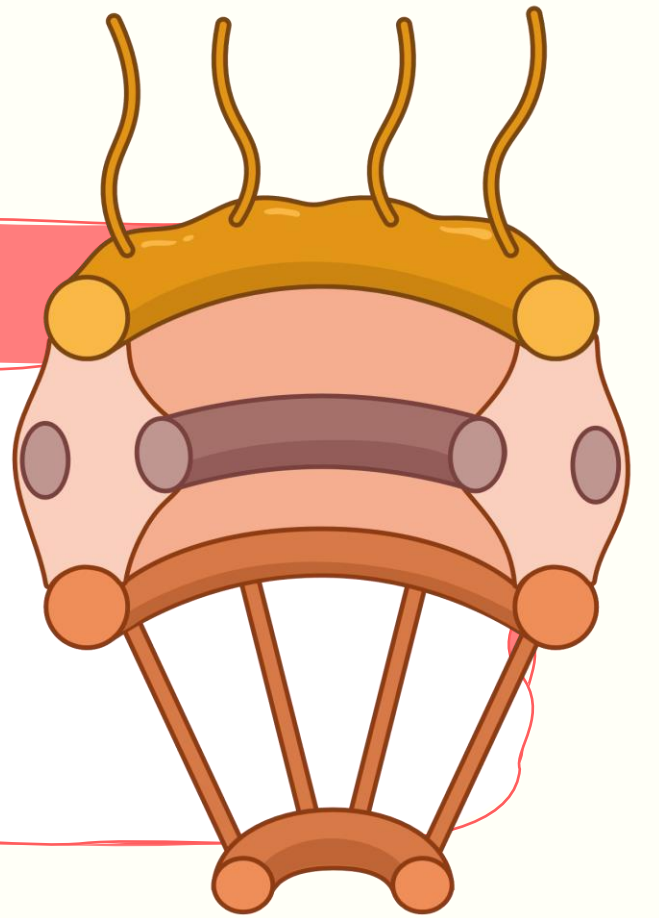
Department of Biological Sciences, University of Bergen, Bergen, N-5020, Norway



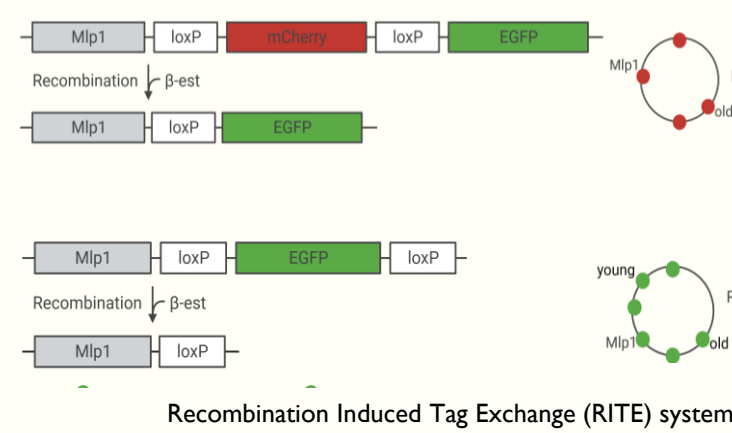
Forskingsrådet
The Research Council of Norway

Abstract

The Nuclear Pore Complex (NPC) is one the largest protein assemblies in the eukaryotic cell, composed of ~1000 proteins. It is responsible all the nucleocytoplasmic communication. Due to the size and complexity, the NPC undergoes a long maturation process. In budding yeast NPC maturation culminates by very late recruitment of MlpI protein. Here we used a combination of inducible protein degradation, fluorescence protein tag exchange and quantitative fluorescence microscopy to investigate the molecular mechanism of late MlpI recruitment.

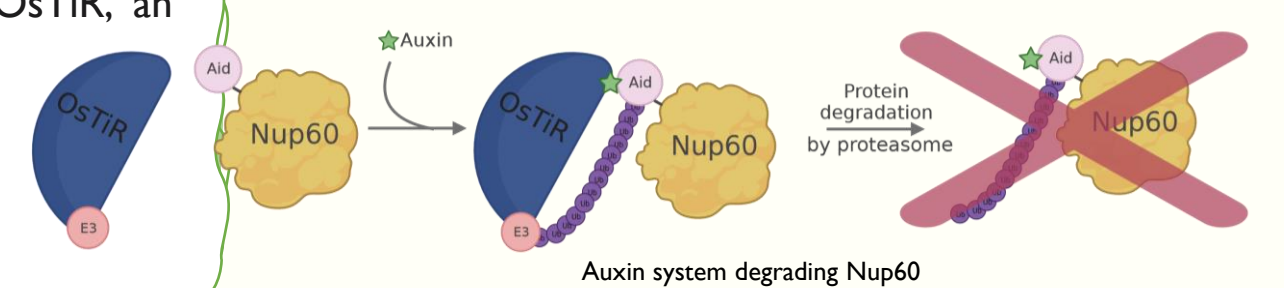


Methods



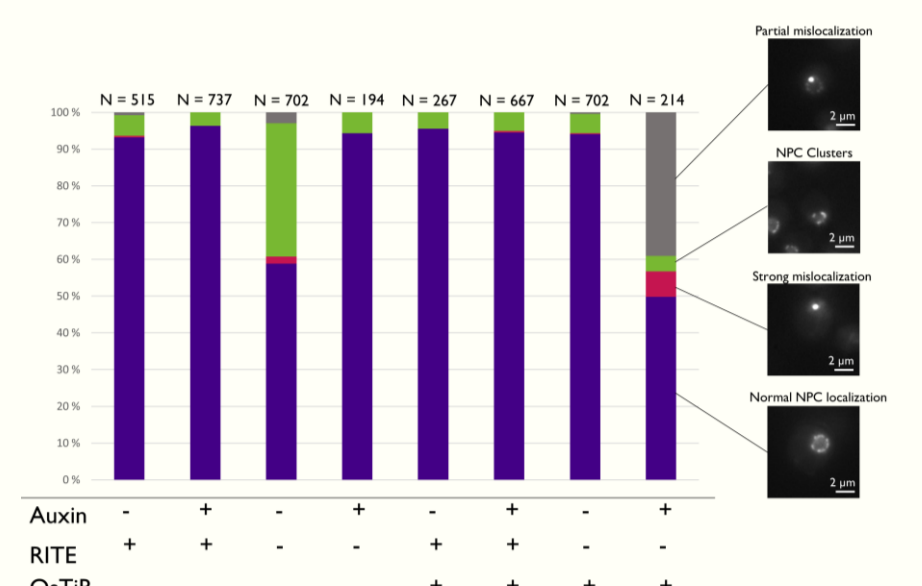
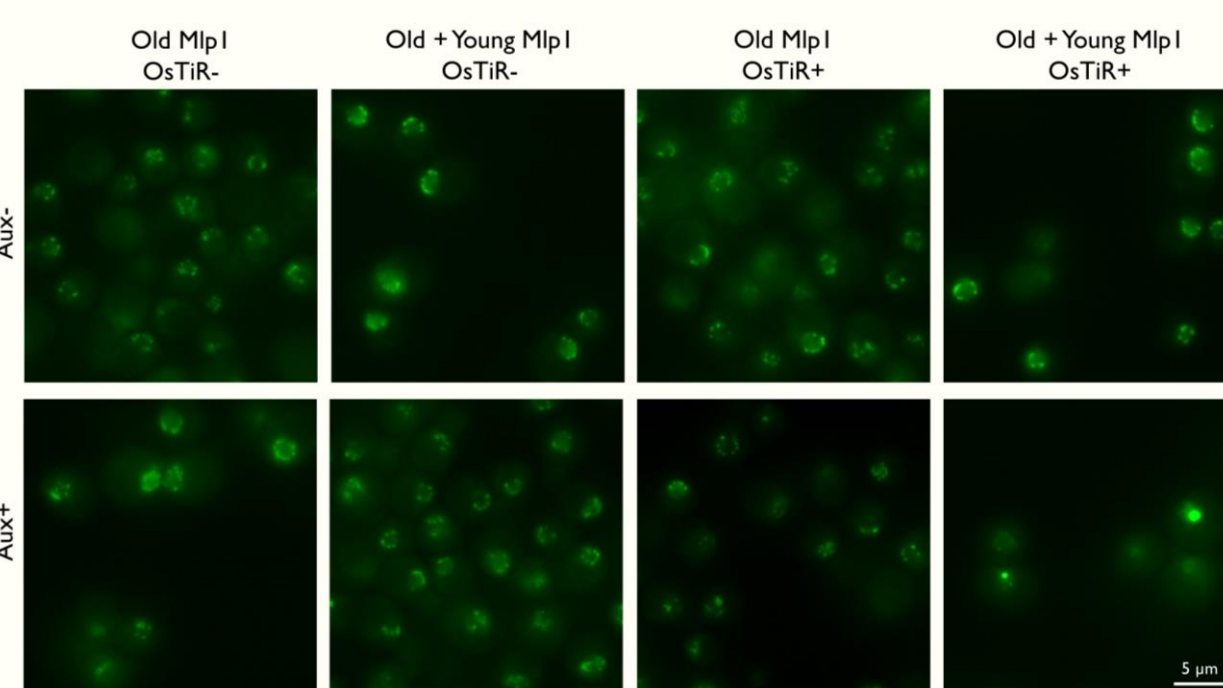
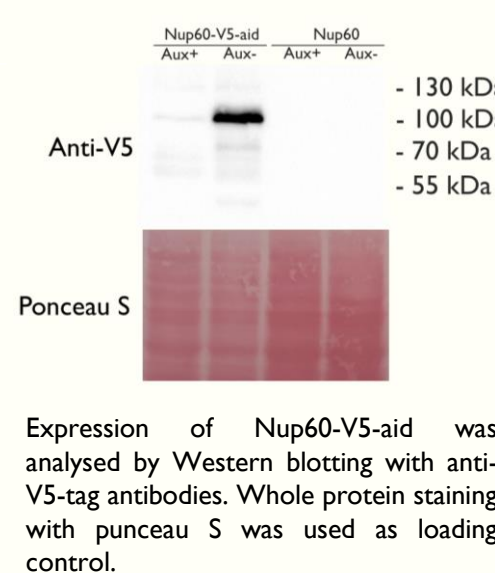
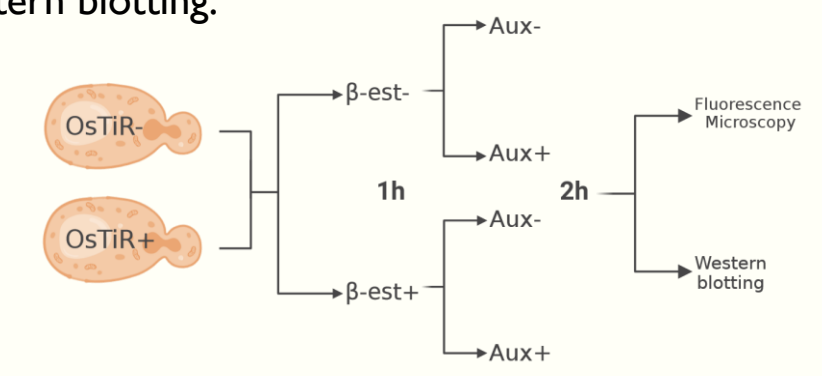
The hormone β -estradiol is added to yeast cultures to induce Recombination Induced Tag Exchange (RITE) system. β -estradiol activates Cre recombinase, which loops out the fluorescent protein tag OTF between LoxP sites. The post-recombination genes either have a second tag outside of the LoxP sites or no tag.

Degradation of Nup60 is induced by a plant-derived hormone called Auxin. The hormone triggers OsTIR, an element of ubiquitin ligase, binding to the aid (Auxin-inducible degron) attached to Nup60. E3 ligase, a part of OsTIR, ubiquitinates the aid of Nup60. Subsequent polyubiquitination promotes rapid proteosomal degradation of Nup60.



MlpI binds NPC at early and late sites differentially dependent on Nup60

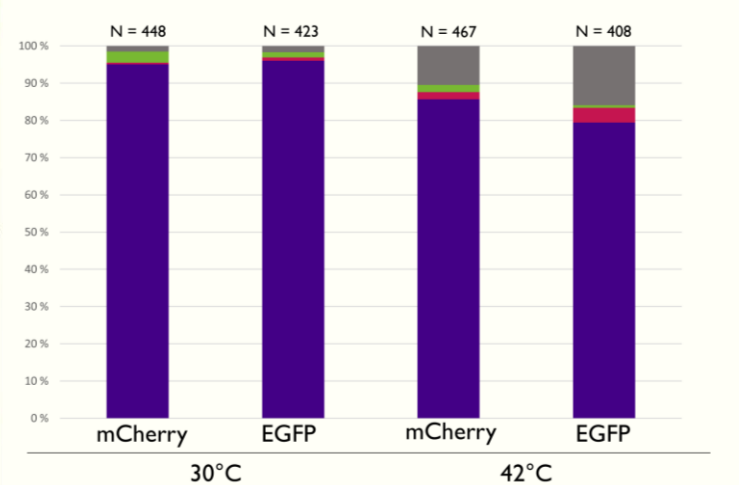
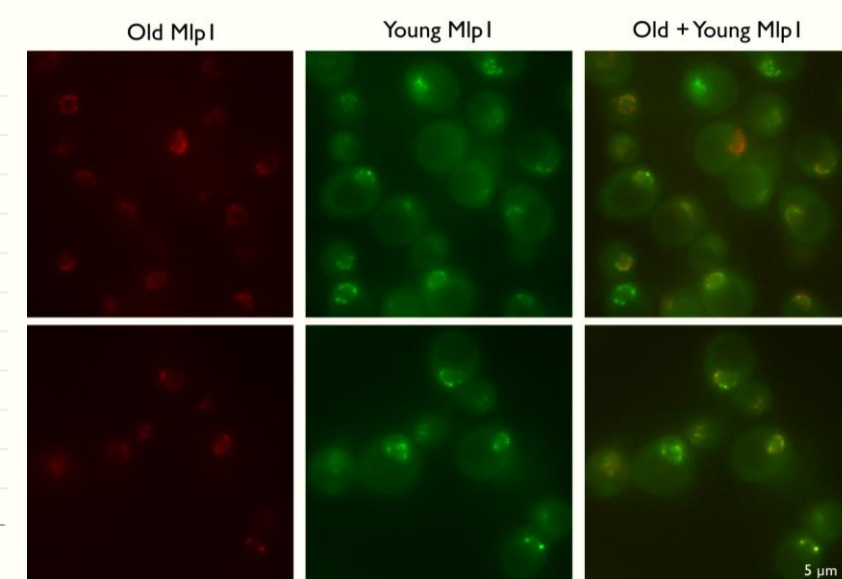
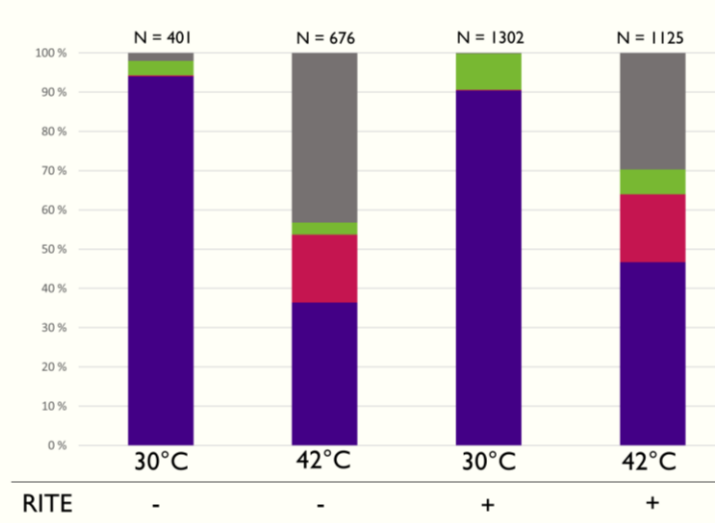
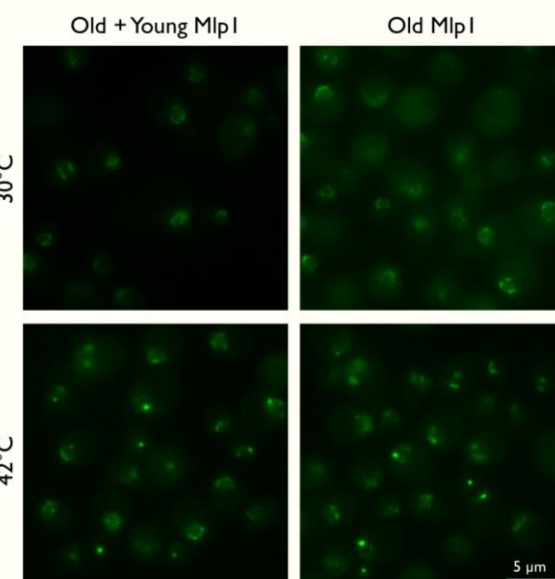
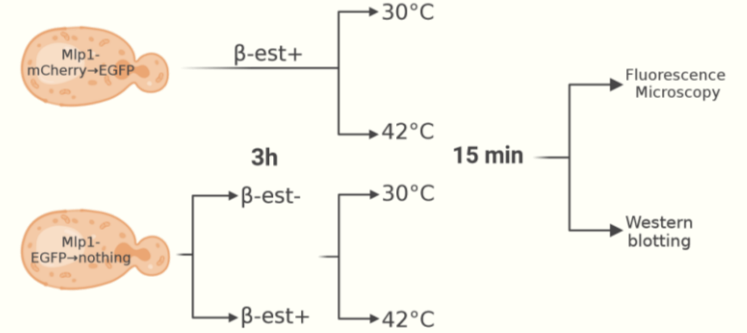
Whole or old population of MlpI was selectively labelled with GFP using recombination induced tag exchange followed by Nup60 degradation via Auxin-dependent degron tag. The yeast cells are then imaged by fluorescence microscopy or analysed by Western blotting.



Representative images and quantification summary of MlpI localization patterns in response to Nup60 depletion

Heat stress disrupts both early and late MlpI binding

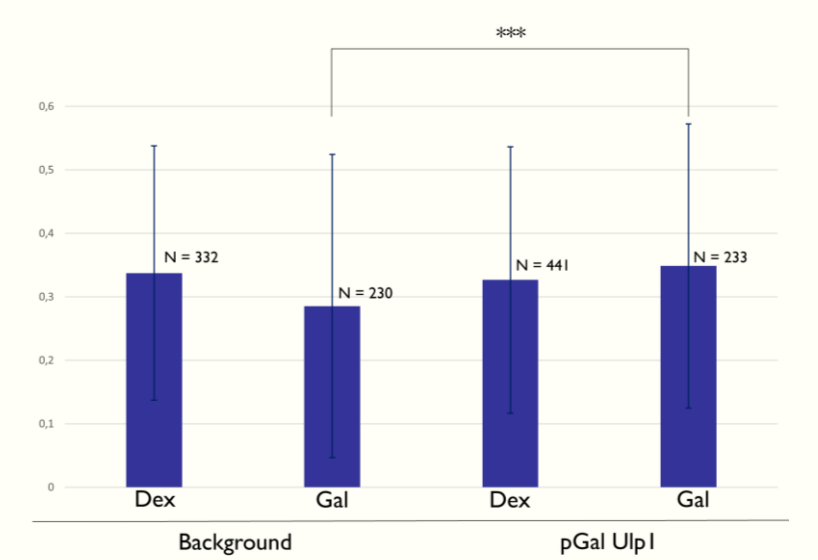
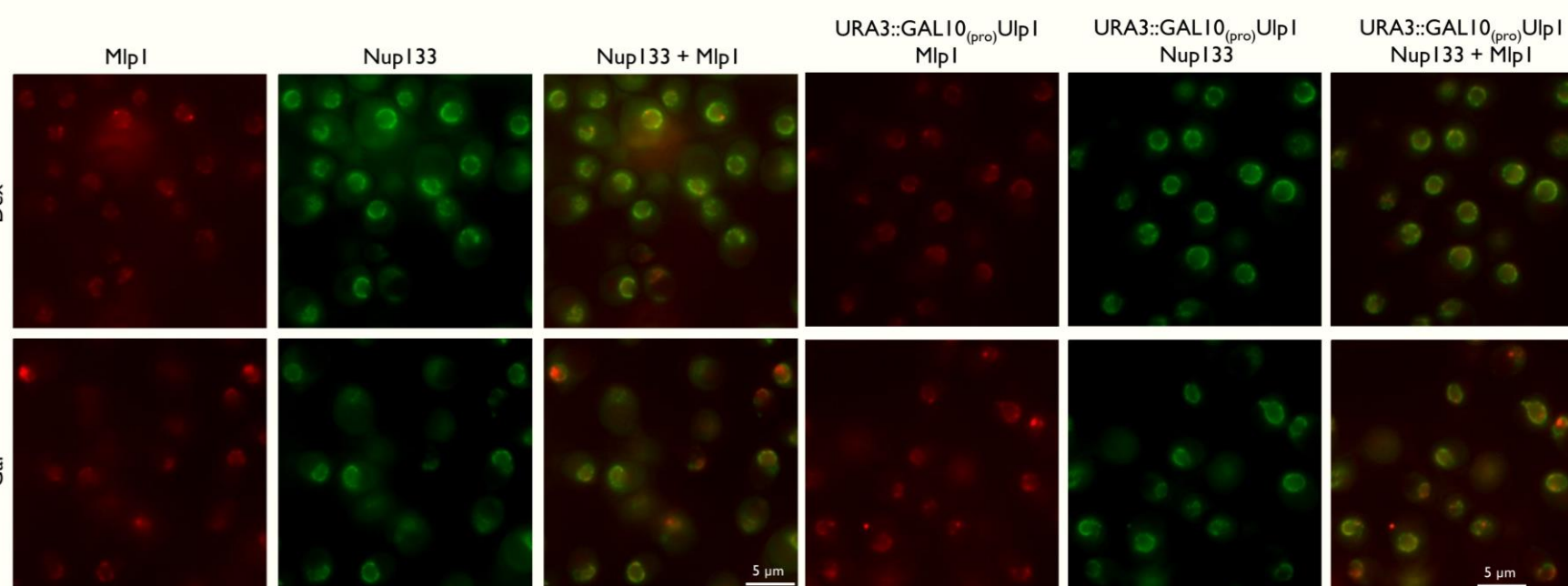
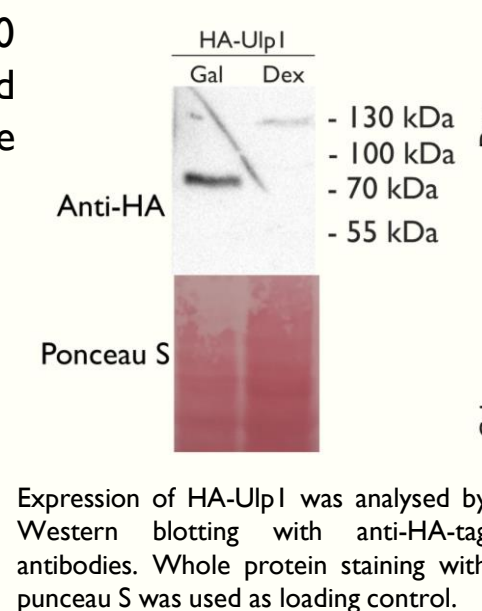
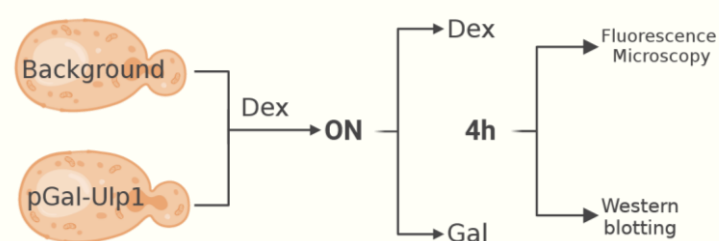
Yeast cells expressing old or young MlpI, selectively tagged with fluorescence protein tags using RITE, were incubated 30°C or 42°C for 15 minutes. MlpI variant localization was analysed by fluorescence microscopy.



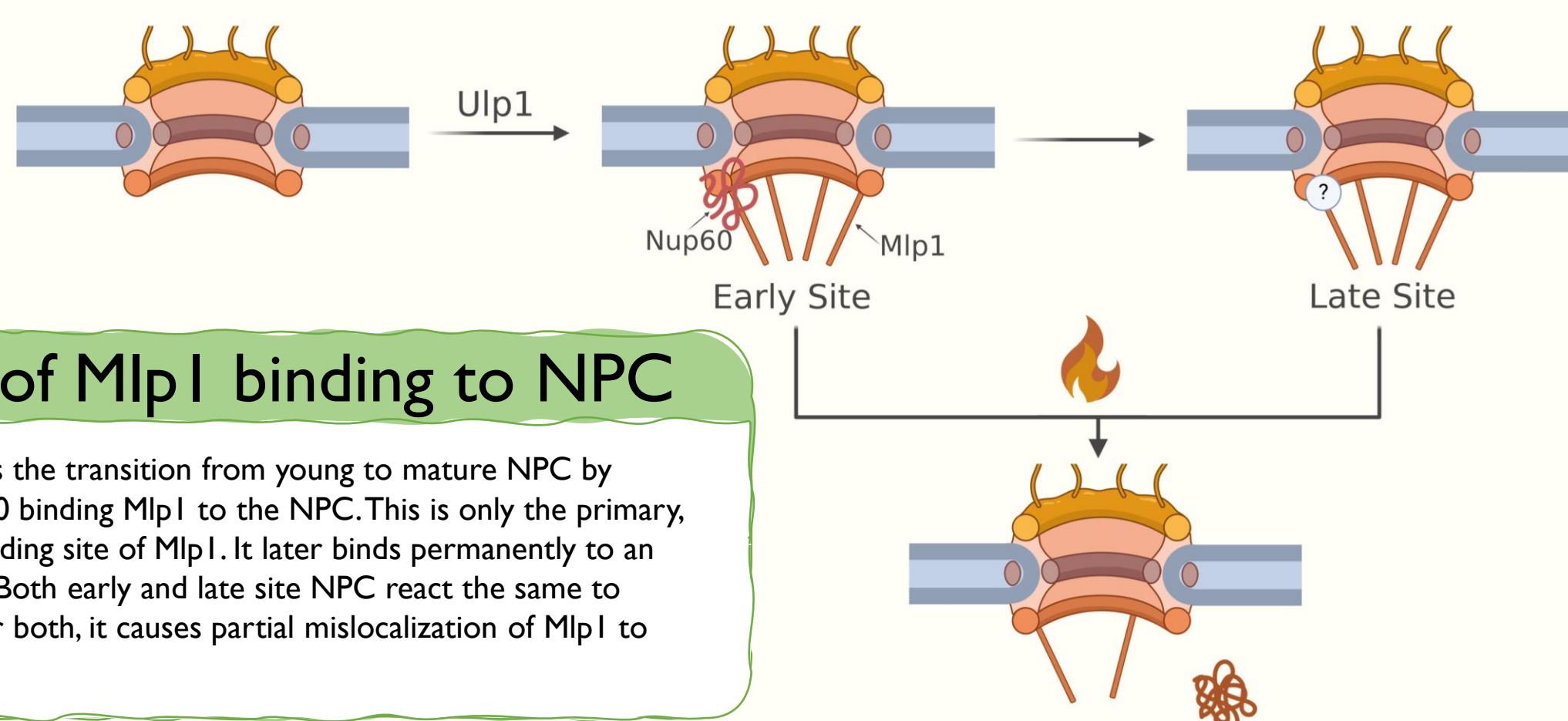
Representative images and quantification summary of MlpI localization patterns in response to heat stress

UlpI may promote MlpI binding to young NPCs

Yeast cells co-expressing MlpI-mCherry, a constitutive NPC marker Nup133-GFP, and UlpI and control of Gal10 promoter in the induced (Galactose medium) or repressed (Dextrose medium) states were imaged by fluorescence microscopy and analysed by Western blotting.



Representative images and quantification summary of MlpI localization patterns in response to UlpI overexpression



Model of MlpI binding to NPC

UlpI promotes the transition from young to mature NPC by assisting Nup60 binding MlpI to the NPC. This is only the primary, transitional binding site of MlpI. It later binds permanently to an unknown site. Both early and late site NPC react the same to heat stress. For both, it causes partial mislocalization of MlpI to the NPC.

References

- Onishchenko, E., Noor, E., Fischer, J. S., Gillet, L., Wojtynek, M., Vallotton, P., & Weis, K. (2020). Maturation Kinetics of a Multiprotein Complex Revealed by Metabolic Labeling. *Cell*, 183(7), 1785–1800.e26. <https://doi.org/10.1016/j.cell.2020.11.001>
- Terweij, M., van Welzen, T., van Deventer, S., Verzijlbergen, K. F., Menendez-Benito, V., Ontoso, D., San-Segundo, P., Neeffjes, J., & van Leeuwen, F. (2013). Recombination-induced tag exchange (RITE) cassette series to monitor protein dynamics in *Saccharomyces cerevisiae*. *G3 (Bethesda, Md.)*, 3(8), 1261–1272. <https://doi.org/10.1534/g3.113.006213>
- Nishimura, K., Fukagawa, T., Takisawa, H., Kakimoto, T., & Kanemaki, M. (2009). An auxin-based degron system for the rapid depletion of proteins in nonplant cells. *Nature methods*, 6(12), 917–922. <https://doi.org/10.1038/nmeth.1401>
- Cibulka, J., Bisaccia, F., Radisavljevic, K., Gudino Carrillo, R. M., & Köhler, A. (2022). Assembly principle of a membrane-anchored nuclear pore basket scaffold. *Science advances*, 8(6), eab6863. <https://doi.org/10.1126/sciadv.ab6863>
- Rajoo, S., Vallotton, P., Onishchenko, E., & Weis, K. (2018). Stoichiometry and compositional plasticity of the yeast nuclear pore complex revealed by quantitative fluorescence microscopy. *Proceedings of the National Academy of Sciences of the United States of America*, 115(17), E3969–E3977. <https://doi.org/10.1073/pnas.1719398115>



SCAN ME